ment channel comprises a single fluid inlet and outlet. Device **70** comprises main electrophoretic flow path **71** in intersecting relationship with secondary electrophoretic flow path **73**. Upstream from the intersection **82** along secondary electrophoretic flow path **73** is enrichment channel **72**. In using this embodiment, sample is introduced through syringe interface **80** into enrichment channel **72**, whereby the analyte comprising fraction of the sample is reversibly bound to the material present in the enrichment channel. An electric field is then applied between electrodes **81** and **79** which moves the non-reversibly bound or waste fraction of the sample out of the enrichment channel **72**, along secondary electrophoretic flow path **73**, past intersection **82**, and out discharge outlet **84** into waste reservoir **78**. An elution buffer is then introduced into enrichment channel **72** through syringe interface **80** and an electric field applied between electrodes **81** and **79**, causing elution buffer to flow through enrichment channel **72** into secondary flow electrophoretic flow path **73**, carrying analyte along with it. When analyte reaches intersection **82**, the electric field between electrodes **79** and **81** is replaced by an electric field between electrodes **76** and **77**, which causes analyte to move along main electrophoretic flow path **71** and towards reservoir **74** through detection region **99**.

The device shown diagrammatically in FIG. **6** comprises an enrichment channel having an electrophoretic enrichment means, instead of the chromatographic enrichment means of the devices of FIGS. **1** to **5**. In device **90**, sample is introduced into reservoir **96** and an electric field is applied between electrodes **87** and **88**, causing the sample to migrate towards reservoir **98**. As the sample migrates towards reservoir **98** it enters stacking gel **93** having a relatively large pore size and travels towards secondary gel **92** of relatively fine pore size. At interface **94**, the sample components are compressed into a narrow band. At this point, the electric field between electrodes **87** and **88** is replaced by an electric field between electrodes **89** and **90**, which causes the narrow band of sample components at interface **93** to migrate into main electrophoretic flowpath **95**, past detection region **91** and towards reservoir **85**. In device **90**, instead of the stacking gel configuration, one could provide for a molecular size membrane at the region of interface **93**, which can provide for selective passage of sample components below a threshold mass and retention at the membrane surface of components in excess of the threshold mass. In yet another modification of the device shown in FIG. **6**, present at the location of interface **93** could be an electrode by which an appropriate electric potential could be applied to maintain a sample component of interest in the region of **93**, thereby providing for component concentration in the region of **93**. For example, for an anionic analyte of interest, upon introduction of sample into reservoir **96** and application of an electric field between **93** and **87**, in which **93** is the positive electrode and **87** the ground, the anionic will migrate towards and concentrate in the region of **93**. After the analyte has concentrated in the region of electrode **93**, an electric field can then be applied between **89** and **90** causing the anionic analyte to migrate towards reservoir **85**.

FIG. **7** provides a top diagrammatic view of a disk shaped embodiment of the subject device, as opposed to the credit card shaped embodiments of FIGS. **3** to **6**. In device **100**, sample is first introduced into enrichment channel **102**. An electric field is then applied between electrodes **108** and **109**, moving elution buffer **103** through enrichment channel **102**, whereby analyte retained in the enrichment channel **102** is released and carried with the elution buffer to intersection **114**. The electric field between **108** and **109** is then replaced with an electric field between **110** and **111**, causing analyte to move from intersection **114** along main electrophoretic flow path **112**, past detection region **113** and towards reservoir **107**.

The subject devices may be used in a variety of electrophoretic applications, where one or more electric fields are applied to a medium to move entities through the medium. Representative electrophoretic applications include separation applications, preparation applications, sequencing applications, synthesis applications, analyte identification applications, including clinical, environmental, quality control applications, and the like. Thus, depending on the particular application a variety of different fluid samples may be introduced into the subject device, where representative samples include bodily fluids, environmental fluid samples, e.g. water and the like, or other fluid samples in which the identification and/or isolation of a particular analyte is desired. Depending on the particular application, a variety of different analytes may be of interest, including drugs, toxins, naturally occurring compounds such as peptides and nucleic acids, proteins, glycoproteins, organic and inorganic ions, steroids, and the like. Of particular interest is the use of the subject devices in clinical applications, where the samples that may be analyzed include blood, urine, plasma, cerebrospinal fluid, tears, nasal or ear discharge, tissue lysate, saliva, ocular scratchings, fine needle biopsies, and the like, where the sample may or may not need to be pretreated, i.e. combined with a solvent to decrease viscosity, decrease ionic strength, or increase solubility or buffer to a specific pH, and the like, prior to introduction into the device. For clinical applications, analytes of interest include anions, cations, small organic molecules including metabolites of drugs or xenobiotics, peptides, proteins, glycoproteins, oligosaccharides, oligonucleotides, DNA, RNA, lipids, steroids, cholesterols, and the like.

The following examples are offered by way of illustration and not by way of limitation.

EXPERIMENTAL

Example 1

High Efficiency Separation of Organic Analytes in an Aqueous Sample

A card as shown in FIG. **4** is used in the separation of organic analytes in an aqueous sample as follows in conjunction with a device that provides for the application of appropriate electric fields through introduction of electrodes into each reservoir of the card and provides for a means of detecting analyte as it passes through detection region **65**. In Card **50**, the enrichment channel **62** comprises porous beads coated with a C-18 phase, while the reservoirs and channels, except for the waste reservoir, comprise 20 millimolar borate buffer. A 100 μl aqueous sample is injected into enrichment channel **62** through interface **66**. Substantially all of the organic analyte in the sample reversibly binds to the C18 coated porous beads, while the remaining sample components flow out of enrichment channel **62** into waste reservoir **63**. 10 μl of an elution buffer (90% methanol/ 10% 20 millimolar borate buffer pH 8.3) are then introduced into the enrichment channel **62** through interface **66**, whereby the reversibly bound organic analyte becomes free in the elution buffer. Because of the small volume of elution buffer employed, the concentration of analyte in the volume of elution buffer as compared to the analyte concentration in the original sample is increased 100 to 1000 times. The seals over reservoirs **57** and **56** are then removed and an electric